55. An Easy and Fast Conversion of N²-[(tert-Butoxy)carbonyl]-L-amino Acids to Corresponding Amino-aldehydes

by Pavol Zlatoidsky

Drug Research Institute, SK-90001 Modra, Slovakia

(15.I.93)

A new method for the preparation of N^2 -[(tert-butoxy)carbony]-L-amino-aldehydes from N^2 -[(tert-butoxy)carbony]-L-amino acids based on reduction of mixed anhydrides with LiAl(t-BuO)₃H is described.

1. Introduction. – N-Protected amino-aldehydes are important intermediates for the preparation of serine and cysteine proteinase inhibitors [1–3] or the formation of pseudo-peptides [4]. Because we are interested in the development of new proteinase inhibitors and because the preparation of some peptide aldehydes was necessary, we developed a fast and straightforward method for the synthesis of N^2 -Boc-amino-aldehydes (Boc = (*tert*-butoxy)carbonyl) as starting material for our subsequent work.

Ito et al. [5] described a synthesis of N^2 -Z-amino-aldehydes (Z-benzyloxycarbonyl; 30–60% yield) via reduction of corresponding protected amino acid methyl esters by Al(i-Bu)₂H. The N^2 -Z-amino aldehydes were purified as semicarbazones because of their high degree of racemisation (by keto-enol tautomerism) on silica gel [5]. These semicarbazones were stable enough during deprotection of common amino-protective groups [5] [10], and they were suitable for the incorporation into the peptide chain and the subsequent deprotection by formaldehyde in acidic medium to give the free aldehyde [5] [6].

Fehrentz and *Castro* [7] described the preparation of very pure N^2 -Boc-amino-aldehydes via LiAlH₄ reduction of N-methoxy-N-methylamides of corresponding acids. The purity of the final products is, according to our experience, determined by the purity of the starting N-methoxy-N-methylamides which were purified by chromatography or by crystallisation.

Argininal and/or its derivatives occupy an exclusive position, and preparation of N^2 -Boc- N^7 -Z-argininal was commonly achieved *via* LiAlH₄ reduction of its δ -lactam [8] [9–11]. The reduction of N^2 -Boc- N^7 -nitroarginin methyl ester by Al(i-Bu)₂H was decribed [5] [12], as well as the LiAlH₄ reduction of *N*-methyl-*N*-methoxyamide [13].

2. Results and Discussion. – We investigated the reduction of mixed anhydrides of Boc-L-amino acids 1 used commonly in peptide chemistry as activating agents, with $LiAl(t-Bu)_3H$. The latter is known as a selective reducing agent for acyl halides [14] and C-acyl formiminium salts [15]. Reduction of N^2 -Boc-L-alanine ethoxycarbonic anhydride (2) with $LiAl(t-Bu_3)H$ yielded a mixture of the desired N^2 -Boc-L-alaninal and ethyl Boc-L-alaninate (3). The formation of 3 is explained by a fast proceeding $S_N i$ substitution due to polarisation of C=O bond by the Li-cation (Scheme 1).



R = Me, i-Pr, Me₂CHCH₂, MeSCH₂CH₂, PhCH₂, (indol-3-yl)methyl

Reduction of mixed anhydrides unable to undergo a rearrangement of the type $2 \rightarrow 3$ should lead to the corresponding aldehydes: Thus, we prepared first the mixed anhydrides 4 derived from diphenylacetic acid because of their easy generation without by-products from N-Boc-L-amino acids and diphenylketene [16] (Scheme 1). The reduction of 4 led to aldehydes 5 contaminated by diphenylacetic acid which was hardly removable. After conversion of the aldehyde 5 to their semicarbazones, the mixtures could be separated by silica-gel chromatography.

Reduction of mixed anhydrides 6, obtained from 1 and pivalic acid in THF with *N*-methylmorpholine as base, yielded the aldehydes 5 in ca. 90% chemical purity (impurities on TLC) with negligible racemisation (according to 'H-NMR; *Scheme 2*).

The accompanying pivalic acid was removed by extraction with diluted Na_2CO_3 solution and by weakly basic resin, and the aldehydes were converted to the semicarbazones 7 [5].



Reaction of N^2 -Boc- N^7 -nitro-L-arginine pivalic anhydride (8) with LiAl(*t*-BuO)₃H yielded cylic lactam 9, known as a by-product in mixed-anhydride-method activation of N^2 , N^7 -diprotected arginines [8]. This observation is in good agreement with the report [17] on the formation of 9 in the reaction of N^2 -Boc- N^7 -nitroarginine isobutoxycarbonic anhydride and organolithium compounds. A fast cyclisation of mixed anhydride 8 due to polarisation of the carbonyl O-atom is proposed to explain the formation of 9. The latter might be converted to the desired aldehyde *via* LiAlH₄ reduction [10].

Experimental Part

1. General. Boc-Amino acids were purchased from *Fluka*, others from *Merck*. TLC: *Silufol UV 254 (CSFR)*; detection by UV light or by spraying with 1% dinitrophenylhydrazine in 2N HCl; mobil phases (ν/ν) : *A*, hexane/AcOEt 1:1; *B*, hexane/AcOEt 2:1; *C*, AcOEt/EtOH 8:1; *D* CHCl₃/EtOH 10:1. NMR Spectra: *Varian-200* spectrometer; in CDCl₃; δ in ppm tel. to Me₄Si as internal standard (= 0 ppm). Optical rotation: at 20° in MeOH (*c* = 1); *Polamat A*. Elemental analyses: *Carlo Erba 1106*.

2. General Procedure for the Pivaloyl Chloride Method. To a stirred soln. of N^2 -Boc-L-amino acid (0.01 mol) and N-methylmorpholine (0.01 mol) cooled to -10° under dry N₂, pivaloyl chloride (0.01 mol) was added. After stirring at -10° for 20 min, the precipitated N-methylmorpholine hydrochloride was removed and the soln. of anhydride **6** filtered under dry N₂ into a pre-cooled flask (CO₂/EtOH bath) through a cooled frite. The soln. of **6** was cooled to -70° and LiAl(*t*-BuO)₃H (0.011 mol, 2.6 g) in THF (20 ml) added dropwise. After the addition the mixture was stirred for 10 min at -70° , poured into 20% citric acid (20 ml), and extracted with AcOEt (4×). The org. layer was washed with 10% Na₂CO₃ (2 × 20 ml), dried (MgSO₄), stirred 10 min with Amberlite IRA-93 (10 g), and evaporated. Volatile by-products were removed at 0.6 Torr.

The removal of precipitated N-methylmorpholine hydrochloride could be omitted, when 2 equiv. of reducing agent were used.

Using N^2 -Boc- N^8 -nitro-L-arginine, lactam 9 was obtained and purified by flash chromatography.

3. Data of Aldehydes 5. Prepared According to Exper. 2. N²[(tert-Butoxy)carbonyl]-L-alaninal: Yield 76%. TLC: $R_{\Gamma}0.55(A)$, 0.36 (B). [α]_D = -25.52. ¹H-NMR: 9.58 (s, 1 H); 5.5 (s, NH); 4.2 (g, 1 H); 3.7 (g, 1 H); 1.45 (s, 9 H); 1.25 (s, 3 H). ¹³C-NMR: 199.7; 155.4; 79.8; 55.6; 28.6; 27.8.

 $N^{2}-[(tert-Butoxy)carbonyl]-t-valinal: Yield 79\%. TLC: R_{f} 0.59 (A), 0.40 (B). [\alpha]_{D} = -11.6. {}^{1}H-NMR: 9.6 (s, 1 H); 5.33 (d, NH); 4.23 (dd, 1 H); 2.29 (m, 1 H); 1.46 (s, 9 H); 0.95 (2d, 6 H). {}^{13}C-NMR: 200.4; 172.2; 81.1; 56.5; 31.2; 24.3; 18.5; 17.9.$

N²-[(tert-Butoxy)carbonyl]-L-leucinal: Yield 85%. TLC: $R_{\rm f}$ 0.68 (*A*), 0.49 (*B*). [α]_D = -30.62. ¹H-NMR: 9.55 (*s*, 1 H); 5.45 (*d*, NH); 4.2 (*m*, 1 H); 1.9–1.45 (*m*, 2 H); 1.45 (*m*, 9 H); 0.95 (*d*, 6 H). ¹³C-NMR: 200.4; 184.2; 80.1; 41.3; 28.5; 24.6; 22.3; 22.1

N²-[(tert-Butoxy)carbonyl]-L-methioninal: Yield 82%. TLC: R_{f} 0.77 (A), 0.56 (B). [α]_D = −21.30. ¹H-NMR: 9.6 (s, 1 H); 5.78 (d, 1 H); 4.3 (m, 1 H); 3.2 (m, 2 H); 2.55 (m, 2 H); 2.1 (s, 3 H); 1.5 (s, 9 H). ¹³C-NMR: 199.7; 175.3; 80.5; 49.4; 33.6; 30.9; 28.1; 20.3.

N²-[(tert-Butoxy)carbonyl]-L-phenylalaninal: Yield 78%. TLC: R_f 0.86 (A), 0.76 (B), [α]_D = −36.03. ¹H-NMR: 9.4 (s, 1 H); 7.25 (m, 5 H); 5.1 (s, NH); 4.2 (m, 1 H); 3.65 (q, 1 H); 3.6 (d, 2 H); 1.45 (s, 9 H). ¹³C-NMR: 199.4; 161.5; 136.6; 129.4; 128.4; 126.6; 58.3; 36.2; 28.2.

N²-[(tert-Butoxy)carbonyl]-L-tryptophanal: Yield 81%. TLC: R_f 0.90 (A), 0.76 (B). $[\alpha]_D = -8.50$. ¹H-NMR: 9.7 (s, 1 H); 7.5–7.6 (d, 1 H); 6.8–7.4 (m, 4 H); 8.3 (s, NH(ind)); 5.3 (s, 1 H); 3.6–3.8 (m, 1 H); 1.5–1.6 (d, 2 H); 1.3–1.45 (s, 9 H). ¹³C-NMR: 200.6; 177.7; 135.5; 127.5; 124.1; 121.7; 120.5; 102.1; 79.3; 55.7; 31.0; 28.7.

N²-[(tert-Butoxy)carbonyl]-N⁷-nitro-L-arginine 1,4-Lactam: Yield 85%. TLC: $R_{\rm f}$ 0.50 (C), 0.79 (D). [α]_D = -2.20. ¹H-NMR: 5.56 (m, 1 H); 4.30 (m, 2 H); 3.80 (m, 2 H); 1.35 (s, 9 H); 1.3 (q, 2 H). ¹³C-NMR: 177.0; 159.7; 156.5; 80.3; 52.5; 42.8; 28.2; 24.9; 24.8.

4. Semicarbazones 7. Semicarbazones were prepared according to [5] and purified by flash chromatography (silica gel (3 × 50 cm), Et₂O (500 ml), then Et₂O/EtOH 6:1 (700 ml)); product detection using a *Büchi* UV spectrometer ($\lambda = 254$ nm).

 N^2 -*[(tert-Butoxy)carbonyl]*-*L-valinal Semicarbazone*: Yield 63%. TLC: R_f 0.46 (*C*), 0.39 (*D*). [α]_D = -14.63. ¹³C-NMR: 176.4; 158.8; 144.2; 80.0; 56.7; 47.1; 31.3; 28.6; 18.4; 18.1. Anal. calc. for C₁₁H₂₁N₄O₃ (258.12): C 50.76, H 8.46, N 21.53; found: C 50.54, H 8.39, N 21.67.

 N^2 -[(tert-Butoxy)carbonyl]-L-leucinal Semicarbazone: Yield 67%. TLC: R_{Γ} 0.57 (C), 0.42 (D). [α]_D = -13.61. ¹³C-NMR: 177.2; 155.7; 139.8; 79.9; 40.6; 28.6; 24.7; 22.9; 14.4; 13.9. Anal. calc. for C₁₂H₂₄N₄O₃ (272.09): C 52.93, H 8.83, N 17.64; found: C 53.10, H 8.89, N 17.81.

 N^{2} -[(tert-Butoxy)carbonyl]-L-methioninal Semicarbazone: Yield 62%. TLC: R_{f} 0.68 (C), 0.44 (D). [α]_D = -12.76. ¹³C-NMR: 176.3; 158.2; 143.5; 79.8; 51.5; 32.6; 30.0; 28.3; 15.5. Anal. calc. for C₁₁H₂₂N₄O₃S (289.95): C 45.51, H 7.58, N 19.31; found: C 45.60, H 7.75, N 19.42.

 $N^{2}-f(\text{tert-Butoxy})$ carbonyl]-L-phenylalaninal Semicarbazone: Yield 64%. TLC: R_{f} 0.72 (C), 0.51 (D). [α]_D = -5.10. ¹³C-NMR: 175.8; 158.6; 143.9; 136.6; 129.4; 128.7; 126.6; 80.5; 53.8; 39.9; 28.2. Anal. calc. for C₁(H₂₁N₄O₃ (305.04): C 59.01, H 6.88, N 13.77; found: C 59.21, H 6.71, N 14.01.

 N^{2} -*f*(tert-*Butoxy*)*carbonylJ*-*L*-*tryptophanal* Semicarbazone: Yield 62%. TLC: R_{f} 0.80 (*C*), 0.56 (*D*). [α]_D = -4.25. ¹³C-NMR: 175.4; 155.6; 138.5; 136.1; 127.8; 123.2; 121.9; 119.6; 118.7; 111.2; 109.8; 80.1; 54.3; 28.8; 17.6. Anal. calc. for C₁₇H₂₂N₅O₃ (344.11): C 59.30, H 6.39, N 20.34; found: C 59.78, H 6.23, N 20.67.

5. General Procedure for the Diphenylketene Method. Diphenylketene was prepared according to [18]. To a soln. of Boc-L-amino acid (0.01 mol) in dry THF (20 ml) Et₃N (20 μ l) was added. The mixture was stirred under dry N₂ and cooled to -15.°. Then, diphenylketene (1.94 g, 0.01 mol) in THF (10 ml) was added dropwisc and the temp. maintained at -15°. After 5 min stirring, the mixture was cooled to -40° and LiAl(*t*-BuO)₃H (2.25 g, 0.01 mol) in THF (20 ml) added dropwise at -40°. After the addition, stirring was continued for 10 min and the mixture processed as in *Exper.* 2. Yields of semicarbazone: 51–69%, after chromatography.

REFERENCES

- [1] I. J. Galpin, G. A. Wilby, G. A. Place, R. J. Beynon, Int. J. Pept. Protein Res. 1984, 23, 477.
- [2] K. Kawamura, S. Kondo, K. Maeda, H. Umezawa, Chem. Pharm. Bull. 1969, 17, 1902.
- [3] K. Tatsua, N. Mikami, K. Fujimoto, S. Umezawa, H. Umezawa, T. Aoyagi, J. Antibiot. 1973, 26, 625.
- [4] J.S. Kaltenbronn, J.P. Hudspeth, E.A. Lunney, B.M. Michniewicz, E.D. Nicolaides, J.T. Repine, W.H. Roark, M.A. Stier, F.J. Tinney, P.K.W. Woo, A.D. Essenburg, J. Med. Chem. 1990, 33, 835, and ref. cit. therein.
- [5] A. Ito, R. Takahashi, Y. Baba, Chem. Pharm. Bull. 1975, 23, 3081.
- [6] A. Ito, R. Takahashi, C. Miuna, Y. Baba, Chem. Pharm. Bull. 1975, 23, 3106.
- [7] J.A. Fehrentz, B. Castro, Synthesis 1983, 676.
- [8] M. Bodansky, J. T. Sheenan, Chem. Ind. (London) 1960, 1268.
- [9] G. Borin, G. Chessa, G. Cavagion, F. Marchiori, G. Chessa, W. Muller-Esterl, *Hoppe-Seylers Z. Phys. Chem.* 1981, 362, 163.
- [10] H. Saeki, Y. Shimada, N. Kavaki, B. Shimidzu, E. Ohki, K. Maeda, H. Umezawa, Chem. Pharm. Bull. 1973, 21, 163.
- [11] R. McConnel, J. Y. Lyndall, D. Frizell, C. Ezell, J. Med. Chem. 1993, 36, 1084.
- [12] R. McConnel, G. E. Barnes, F. C. Hozung, J. M. Gunn, J. Med. Chem. 1990, 33, 86.
- [13] A. Murphy, R. Pagnino, Jr., P. L. Vallar, A. J. Tripe, S. L. Sherman, R. H. Lumpkin, S. Y. Tamura, T. R. Webb, J. Am. Chem. Soc. 1992, 114, 3156.
- [14] H.C. Brown, Subba Rao, J. Am. Chem. Soc. 1958, 88, 5372.
- [15] T. Fujisawa, T. Mori, S. Tsuga, T. Sato, Tetrahedron Lett. 1983, 24, 1543.
- [16] G. D. Losse, E. Demuth, Chem. Ber. 1961, 94, 1762.
- [17] J. DiMaio, B. Gibbs, J. Lefebvre, J. Konishi, D. Munn, Shi Yi-Yue, J. Med. Chem. 1992, 35, 3331.
- [18] H. Steudinger, Ber. Dtsch. Chem. Ges. 1911, 44, 1619.